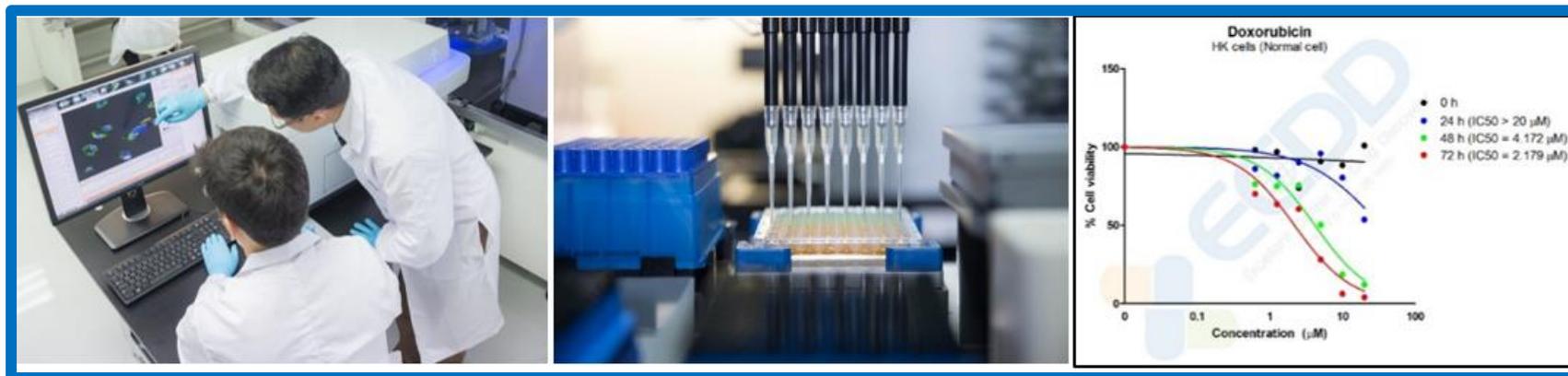


## A Role of ECDD in Leading Natural Products for Drug Discovery and Development in Thailand



**Suparerk Borwornpinyo**, Founder and Director  
Excellent Center for Drug Discovery (ECDD), Mahidol University

13<sup>th</sup> April 2021



Approximately more than 60% of drugs available on the market today are derived from chemical structures found in nature.



Yet over the past few decades, the influence of natural products on drug discovery has notably reduced.



A renewed interest in natural products, the urgent need for new drugs.

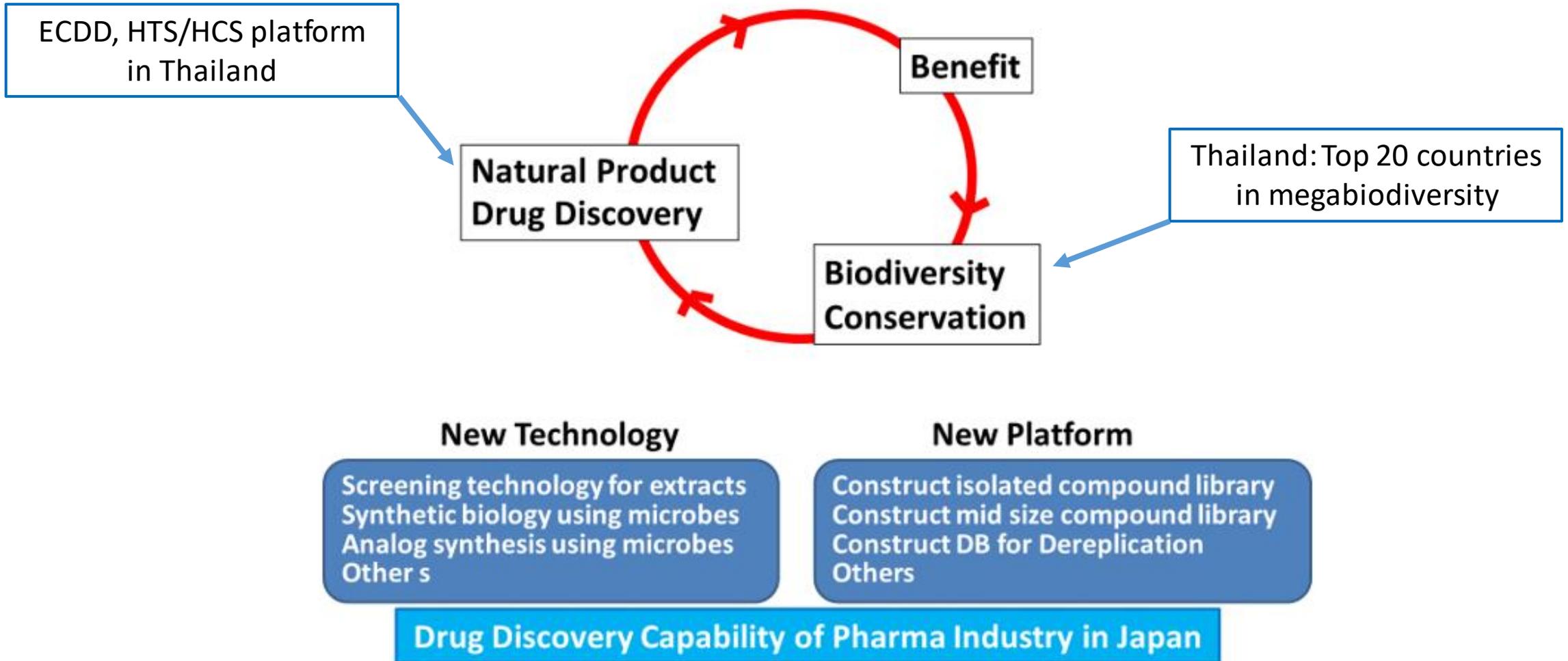
The perceived difficulties of isolating and synthesizing these complex molecules



The challenges associated with screening them using high throughput assays



# NPDD Ecosystem in Asia: An Open Innovation Platform



# APAC DA-EWG: Pillar 5

## Drug Discovery using Natural Product



### Preparation for Pilot Project

#### Internship for technology transfer (Bilateral communication)

- 4 month (February-May, 2019) internship at Takeda Shonan Research Center (tech-transfer & capacity building)
- Mr. Phongthon Kanjanasirirat (ECDD, Mahidol University)



# Overall scheme of the NPDD initiative with Thai institutions

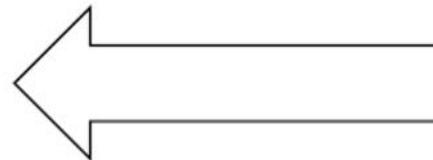
## Capacity building through phenotypic screening

### iPark/Japan



**MN cell death assay**  
**MNP stocks**

1. Tech transfer for HTS/HCS
2. Drug discovery knowledge



**Masaaki-san** **Keiko-san**



1. Receive the results
2. Make decision for further collaboration

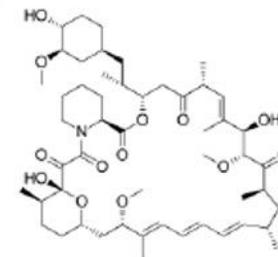
### Thailand



After 3-6 months  
Return to Thailand

**1<sup>st</sup> June**

Perform HTS/HCS for  
Natural Product(NP) lib



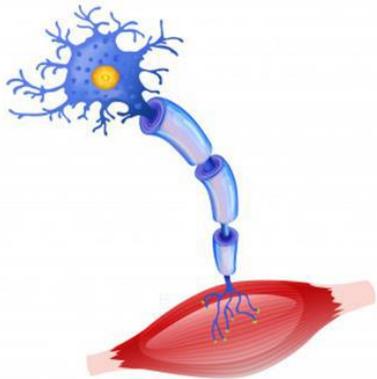
Hit compounds

**August – October 2019**  
**Assay optimization**



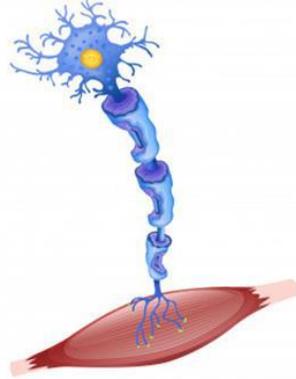
## Amyotrophic Lateral Sclerosis (ALS)

normal nerve cell



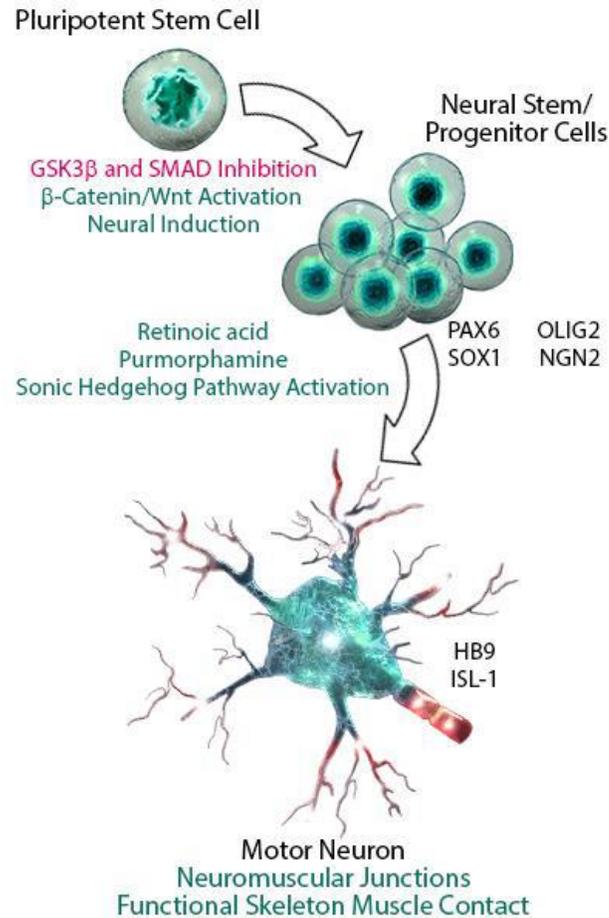
muscle contracts

nerve with sclerosis

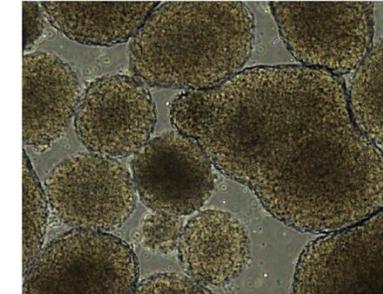


muscle unable to contract

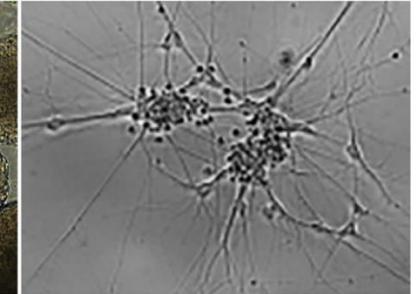
## Motor neuron (MN) deficiency



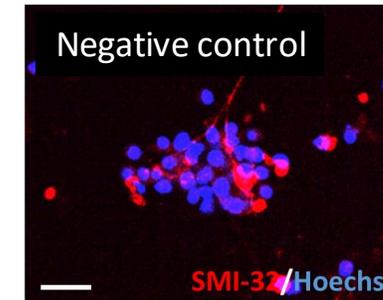
NPC



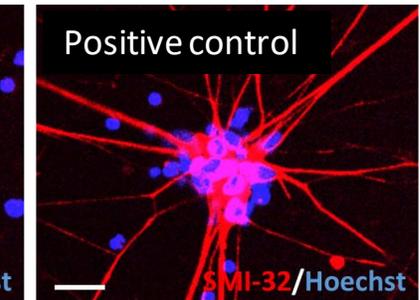
MN cells



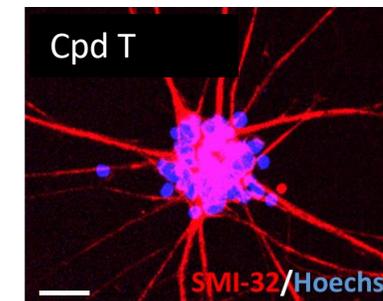
Negative control



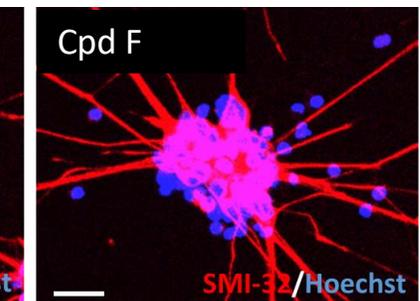
Positive control



Cpd T



Cpd F



Note: SMI-32 = Neurofilament H  
Hoechst = Nucleus  
Scale bar = 25  $\mu$ m





Robotic system

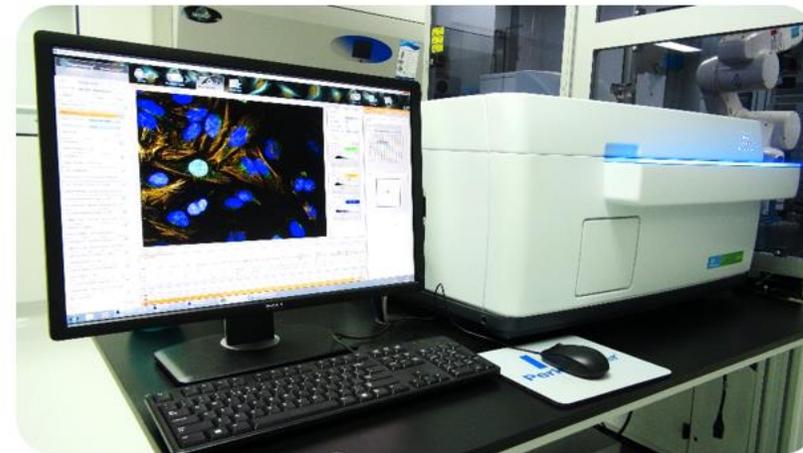
established in 2016:  
TCELS-Mahidol University  
(SC and MD Ramathibodi Hospital)



Compound plate



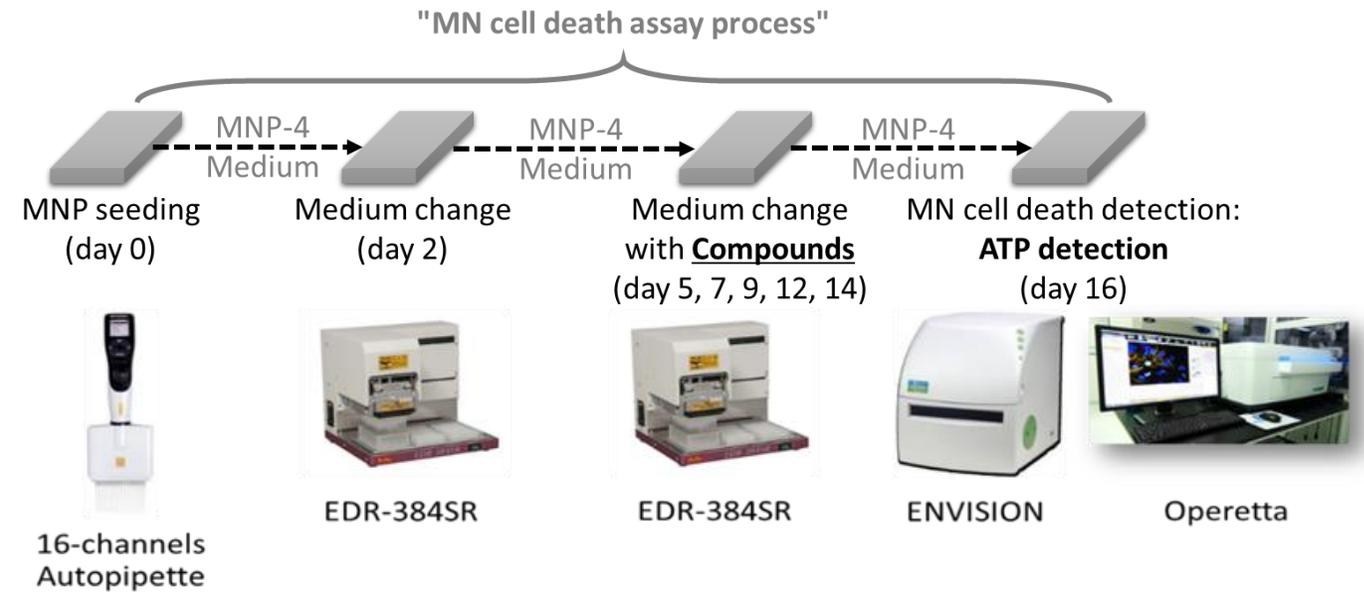
Assay plate (Cell plate)



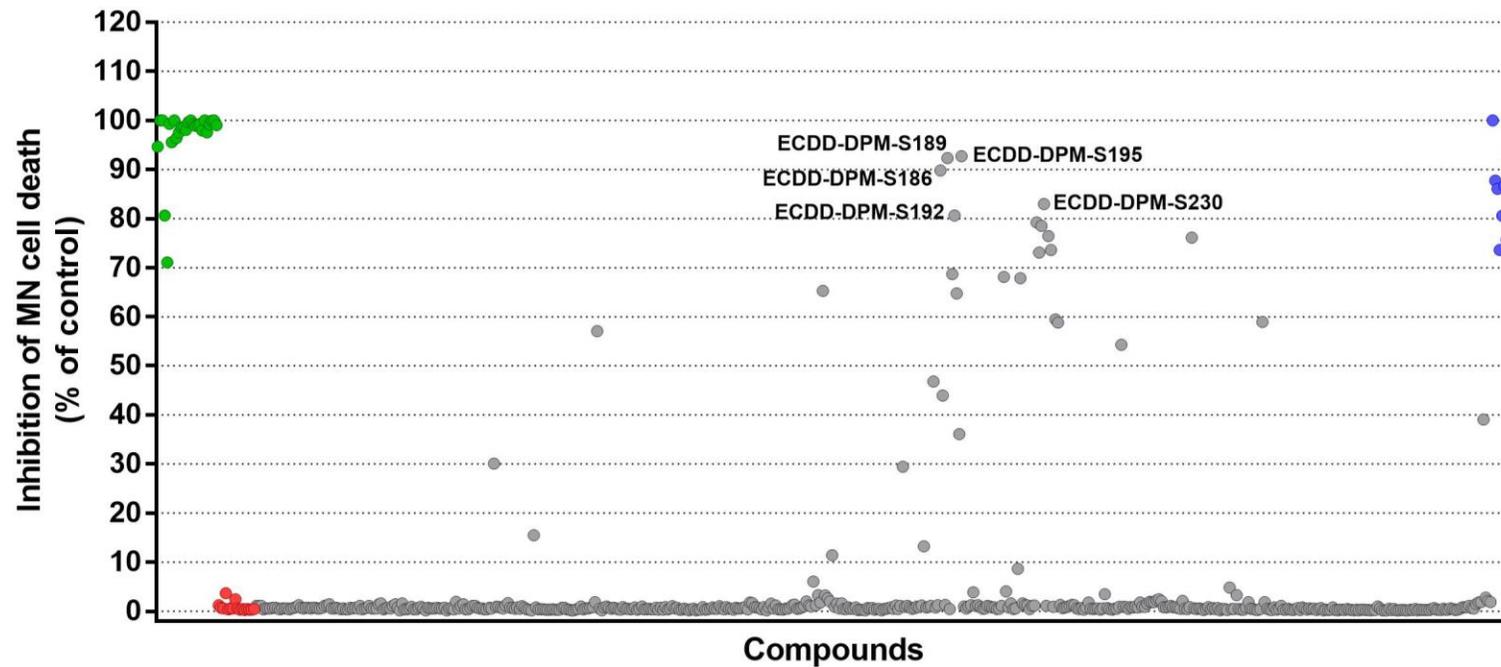
High-Content Imaging system

## Methodology

- Cell numbers: 7,500 cells/well
- Concentration of compounds for screening:
  - Crude extract = 10 µg/ml
  - Purified compound = 10 µM
  - Synthesis compound = 10 µM

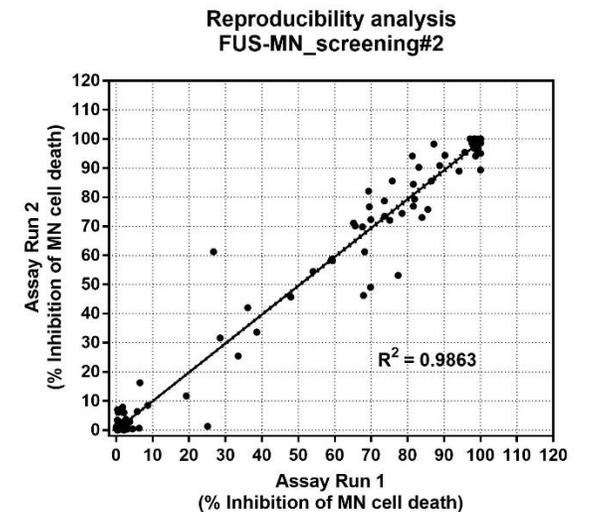


## The Results of an anti-MN cell death screening

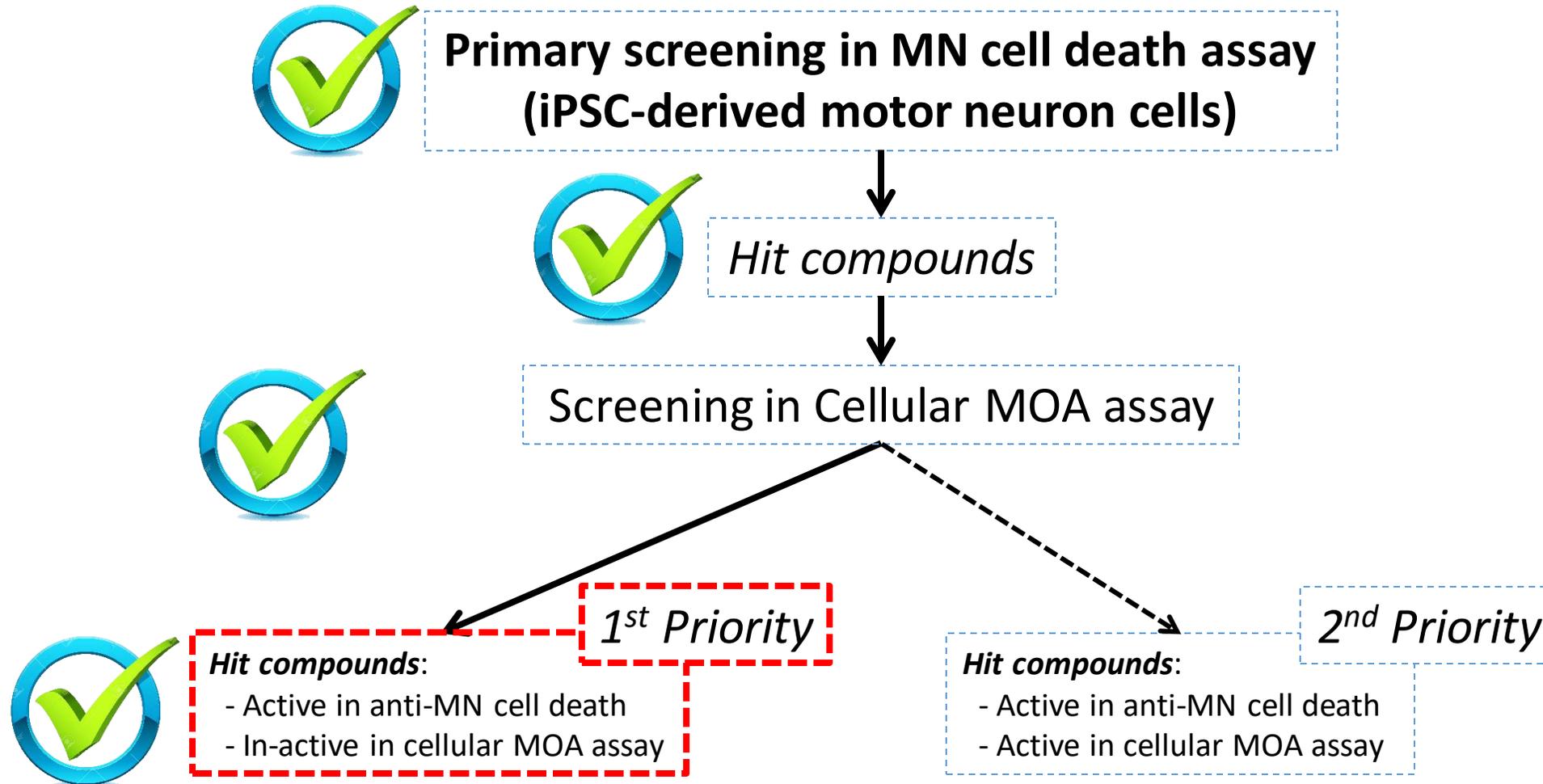


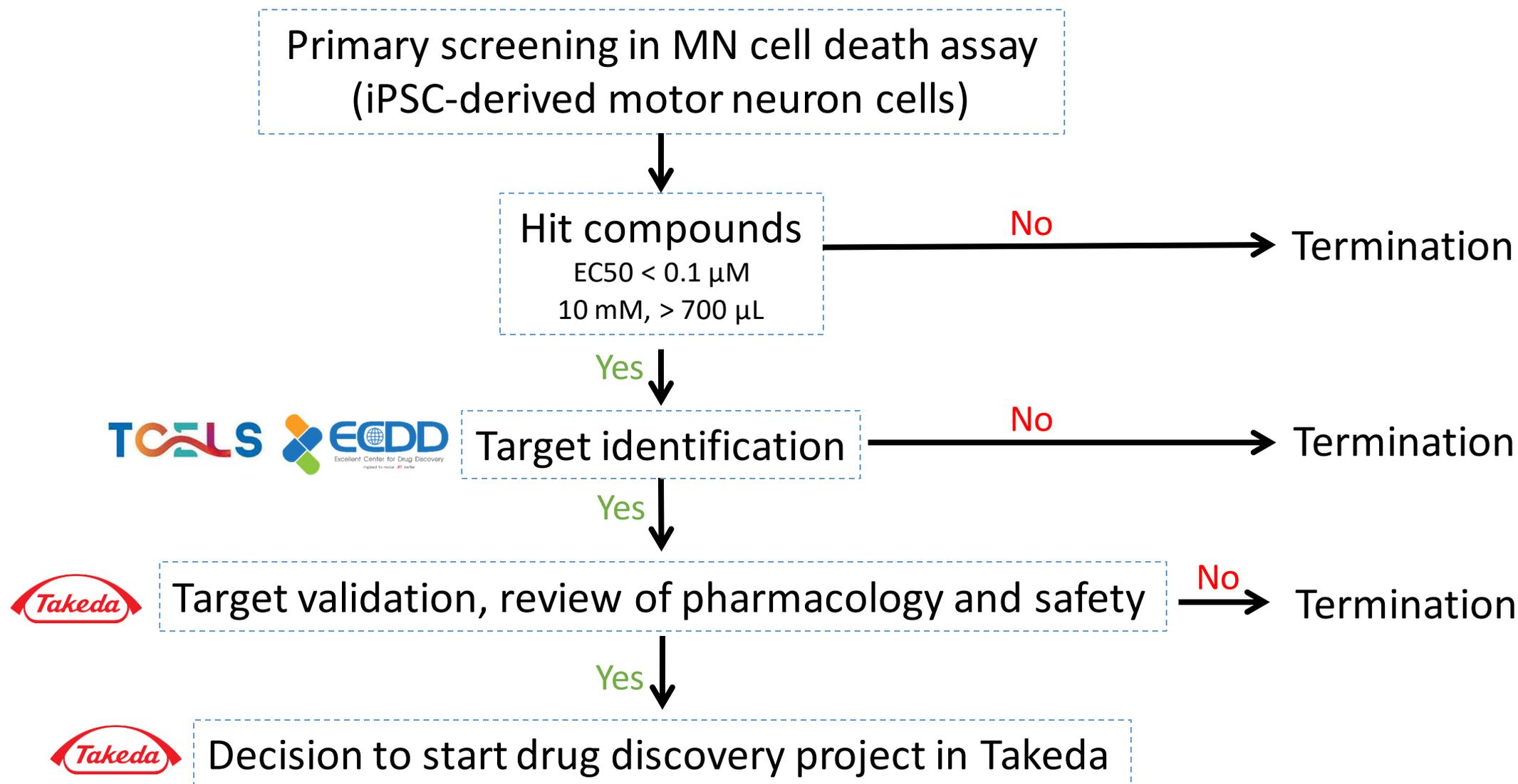
- Positive control
- Negative control
- Compounds
- Ferrostatin-1

**Z-factor = 0.688**



## Screening Process





## ★ First priority selection (IC50 < 0.1 for cellular MOA assay)

No.	Compounds	IC50 MN	IC50 MOA assay
1	ECDD-DPM-N924	6.77	non-effective
2	ECDD-DPM-N937	3.06	non-effective
3	ECDD-DPM-S186	3.06	6.56
4	ECDD-DPM-S189	5.95	4.87
5	ECDD-DPM-S192	non-effective	16.97
6	ECDD-DPM-S195	1.2	4.45
7	ECDD-DPM-S230	3.32	non-effective
8	ECDD-DPM-E971	non-effective	non-effective
9	ECDD-DPM-E1095	5.69	non-effective
10	ECDD-DPM-E1168	1.85	non-effective
11	ECDD-DPM-E1171	0.05	non-effective
12	ECDD-DPM-E1183	1.46	9.26
13	ECDD-DPM-E1204	0.38	non-effective
14	ECDD-DPM-E1211	3.53	non-effective
15	ECDD-DPM-E1215	9.52	non-effective
16	ECDD-DPM-S685	non-effective	7.57
17	ECDD-DPM-S777	3.13	non-effective
18	ECDD-DPM-S443	0.08	3.25
19	ECDD-DPM-S461	0.63	non-effective
20	ECDD-DPM-S465	0.33	non-effective

No.	Compounds	IC50 MN	IC50 MOA assay
21	ECDD-DPM-N131	non-effective	non-effective
22	ECDD-DPM-N190	0.2	1.47
23	ECDD-DPM-N197	3.54	6.11
24	ECDD-DPM-N664	3.54	non-effective
25	ECDD-DPM-N745	0.12	non-effective
26	ECDD-DPM-E278	0.07	0.52
27	ECDD-DPM-E326	0.24	non-effective
28	ECDD-DPM-E329	0.06	non-effective
29	ECDD-DPM-E651	5.15	non-effective
30	ECDD-DPM-E691	10.44	non-effective
31	ECDD-DPM-E707	0.66	non-effective
32	ECDD-DPM-E760	non-effective	non-effective
33	ECDD-DPM-E824	2.91	4.44
34	ECDD-DPM-E825	5.05	non-effective
35	ECDD-DPM-E829	non-effective	non-effective
36	ECDD-DPM-E831	2.86	non-effective
37	ECDD-DPM-E865	non-effective	non-effective
38	ECDD-DPM-E868	7.31	non-effective
39	ECDD-DPM-E869	3.64	non-effective
40	ECDD-DPM-E873	3.51	non-effective



Prof. Wanchai De-Eknamkul



Dr. Lily Eurwilaichitr

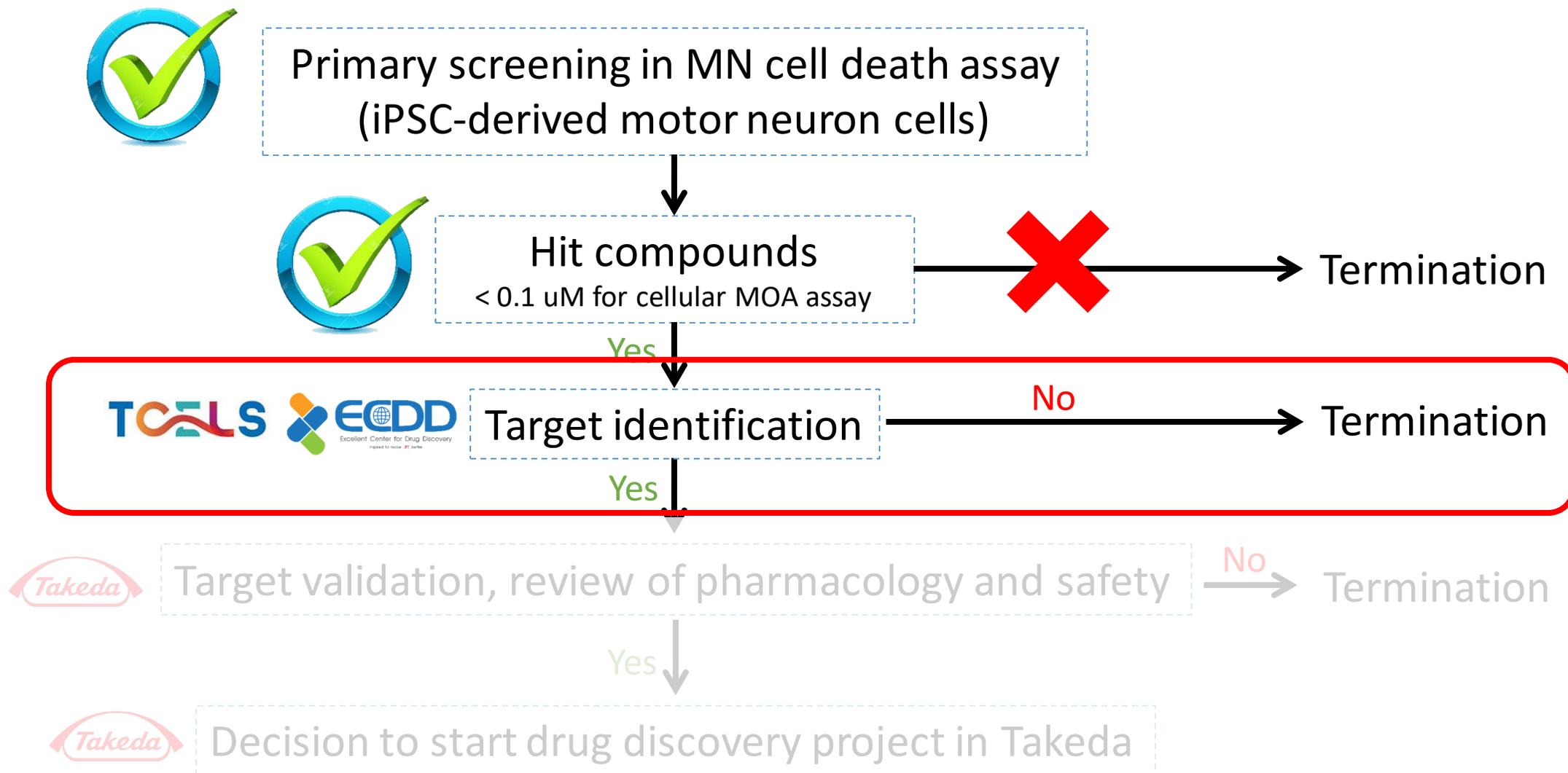


Assoc. Prof. Dr. Surat Laphookhieo



12 Prof. Patoomratana Tuchinda

# Flow of collaboration, decision, plans and actions



**To plan the experiments for  
Target deconvolution**



*Assay optimization & validation*



*Potential targets of hit compounds*

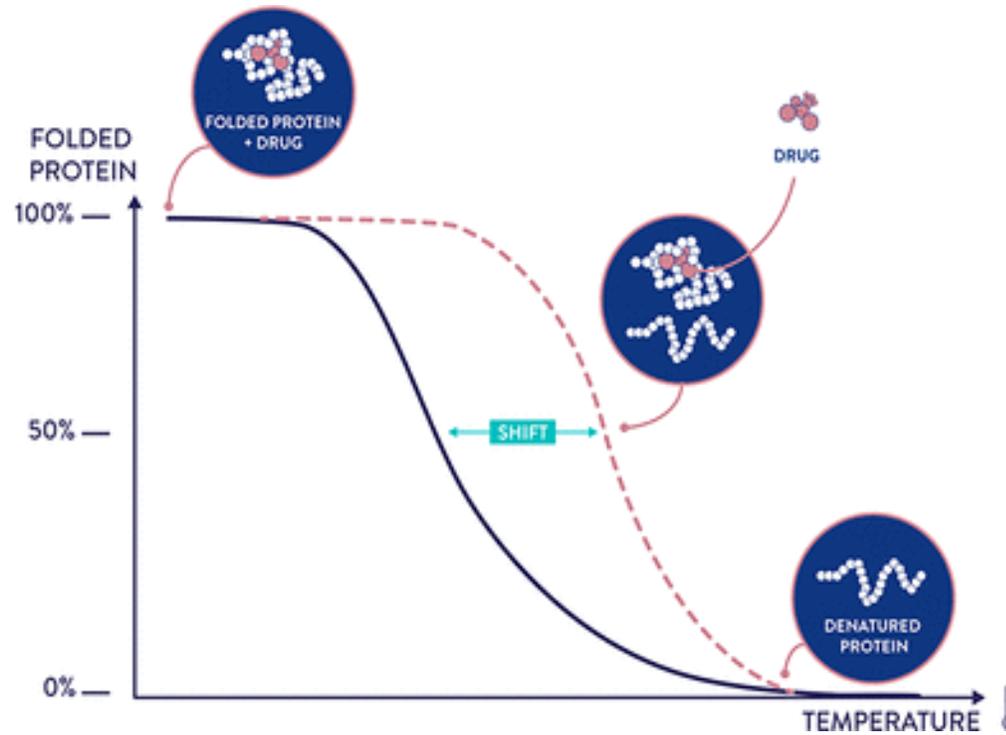


**Dr. Sitthivut Charoensutthivarakul**  
School of Bioinnovation, Mahidol University

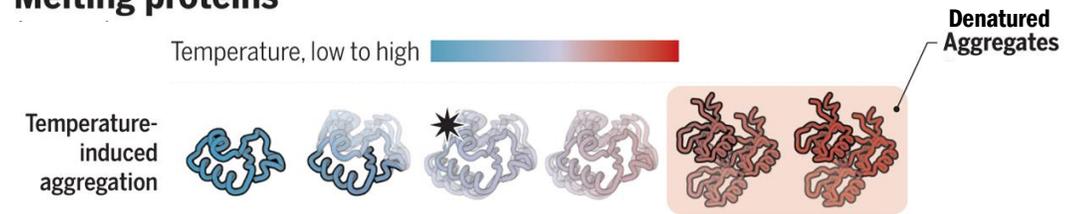


- **Thermal shift assay**

### SAMPLE SOURCES



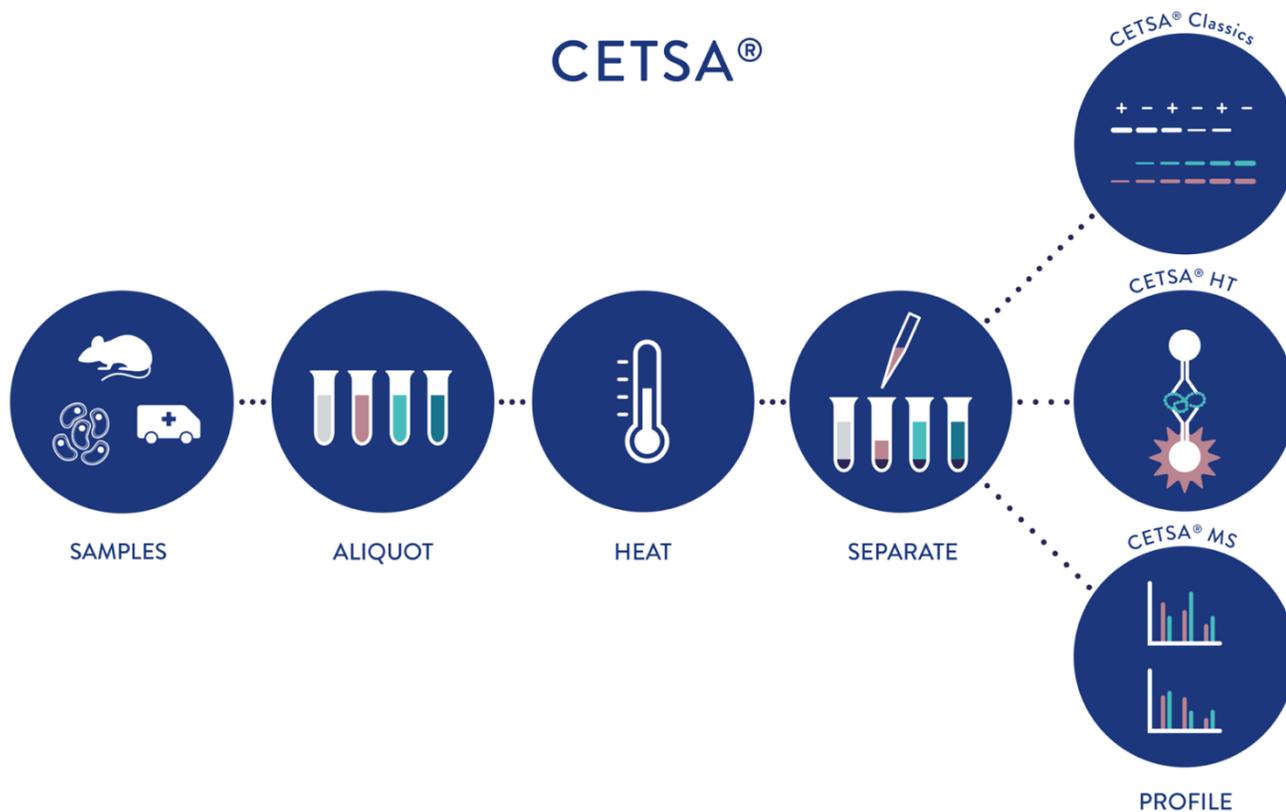
### Melting proteins



A **thermal shift assay (TSA)** measures changes in the thermal denaturation temperature. Upon heating a protein will encounter a temperature at which it denatures, referred to as the melting point. This melting temperature is a physical property and a constant for any given set of conditions. Compounds that interact with a protein will change the melting temperature (thermal shift).



- Experimental workflow



The **CELLular Thermal Shift Assay** is a method that allows the quantification of a compound's target engagement within living cells or in disrupted cells.

The CETSA principle is based on the change in thermal denaturation profile of the target protein that occurs following the binding of a compound. However, in contrast to traditional Thermal Shift Assay, that are carried out in highly purified and isolated systems monitoring a single protein species, CETSA can be performed in complex protein samples and in live cells.

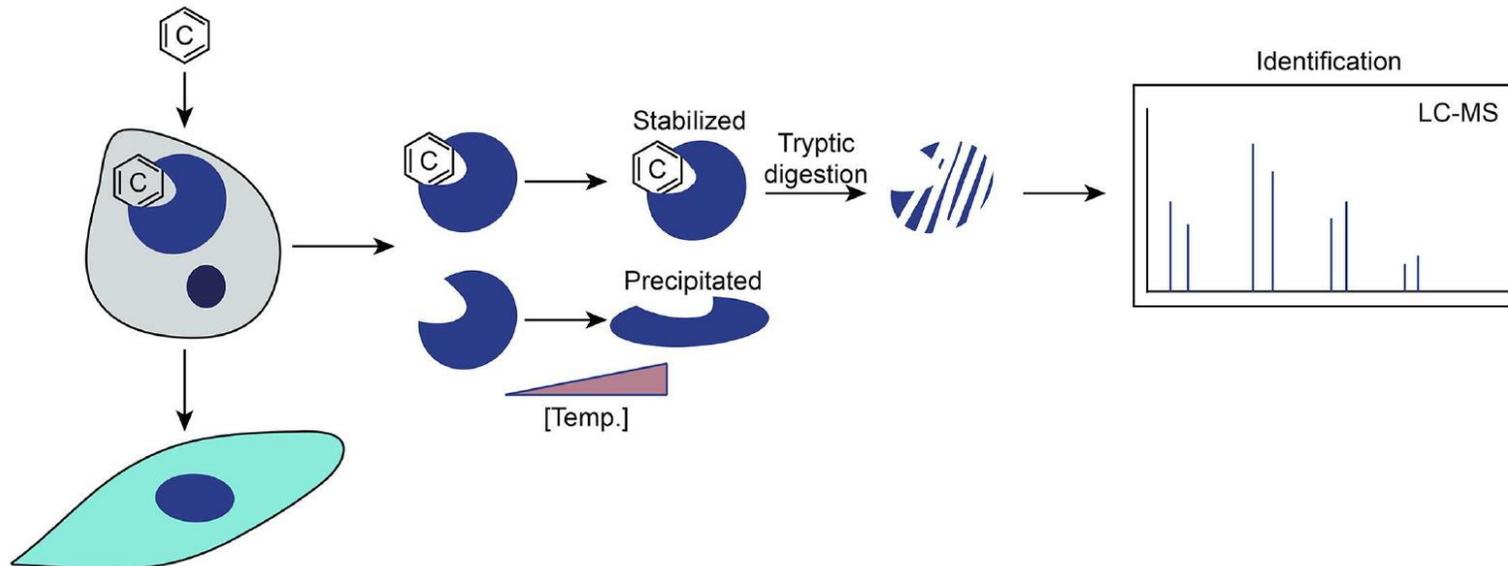
The CETSA is performed by incubating the cells with the test compound, followed by heating of the compound-treated cells, and then by measuring the remaining soluble target protein.

There are three main formats in the CETSA technology platform. Two of the formats, CETSA Classics and CETSA High Throughput (HT) are both targeted CETSA methods for confirming target engagement of a single known protein target using antibodies for the quantification. The third format, CETSA MS, is proteome-wide measurement of cellular target engagement using mass spectrometry.



- CETSA-MS**

MS-based CETSA<sup>®</sup> for target deconvolution in phenotypic drug discovery

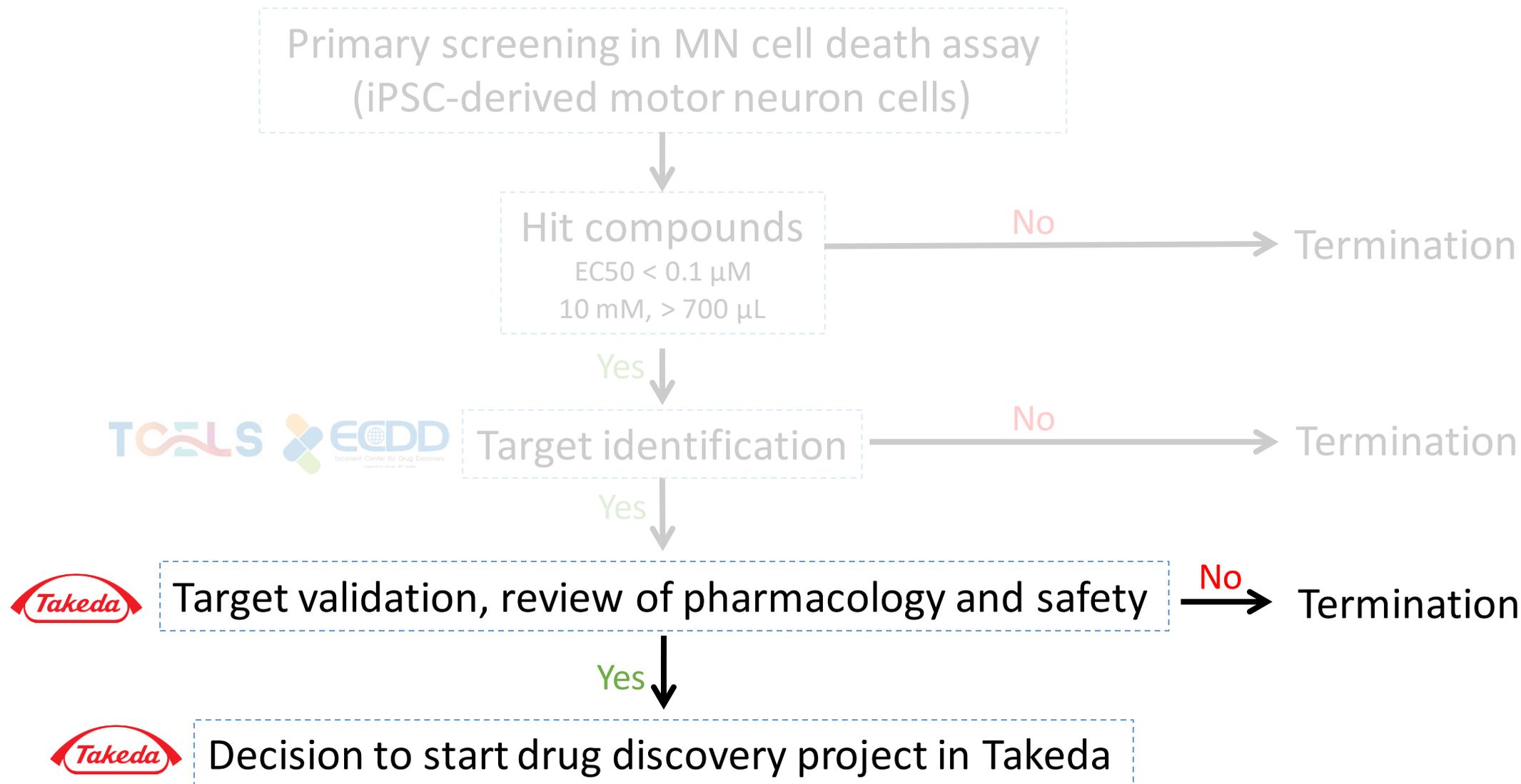


The antibody-based readout enables CETSA to measure the stability shifts of different target proteins in a protein mixture, while this method requires prior knowledge of interested targets. Thus, it is not suitable for unbiased drug target discovery and also cannot be conducted at proteome scale.

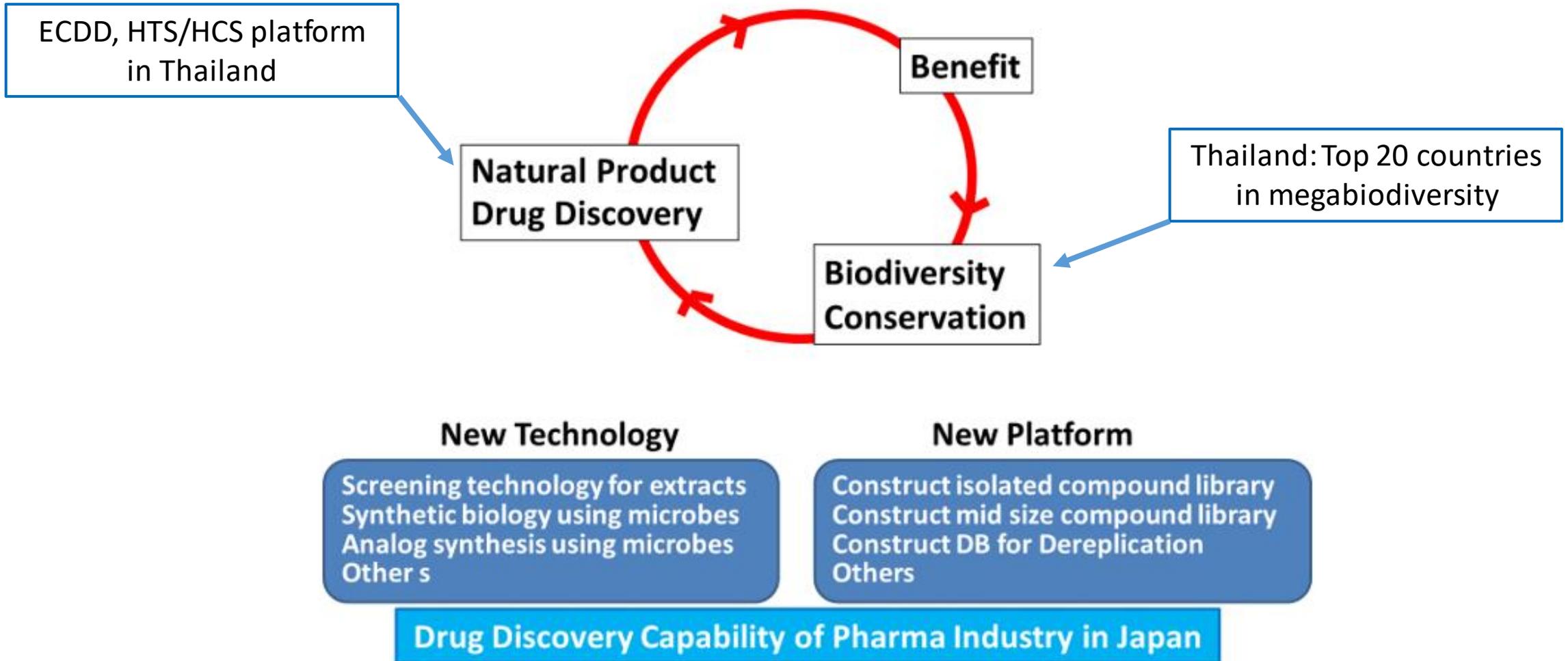
To solve these problems, The Cellular thermal shift assay followed by MS (CETSA-MS) allows for an unbiased search of drug targets and can be applied in living cells without requiring compound labeling.

To date CETSA-MS has been used in several studies for target deconvolution, which have both confirmed previously known compound – target interactions and discovered new ones.





# NPDD Ecosystem in Asia: An Open Innovation Platform



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